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Maus Laboratory

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Using the immune system as a cancer treatment has the potential to induce long-term, durable remissions, and perhaps even cure some patients. The T cells of the immune system are able to specifically kill the target cells they recognize. T cells are also able to persist in the body for many years, and form immune ‘memory,’ which enables the possibility of long-term protection.

The Maus laboratory is interested in using genetic engineering techniques to re-direct T cells to find and kill tumor cells, while sparing healthy tissues. We aim to develop new ways to design molecular receptors to target T cells to liquid and solid tumors; use T cells as delivery vehicles for other drugs, and use drugs to help T cells work against tumors; and understand how T cells can work as “living drugs” to treat patients with cancer.

Immune therapies that engage T cells have the potential to induce long-term durable remissions of cancer. In hematologic malignancies, allogeneic hematopoietic stem cell transplant can be curative, in part due to T-cell mediated anti-tumor immunity. In solid tumors, checkpoint blockades with anti-CTLA-4 or anti-PD-1 monoclonal antibodies can mediate long-term responses by releasing T cells from tightly controlled peripheral tolerance. Our laboratory focuses on T cell biology and T cell engineering. We design chimeric antigen receptors (CARs) to re-direct T cells to specific antigens. Redirecting T cells with CARs is an alternative method of overcoming tolerance and has shown great promise in the clinical setting for B cell malignancies such as leukemia and lymphoma. However, successful application of this form of therapy to other cancers is likely to require refinements in the molecular and clinical technologies.

The goal of the Maus lab is to design and evaluate next generation genetically-modified T cells as immunotherapy in patients with cancer.

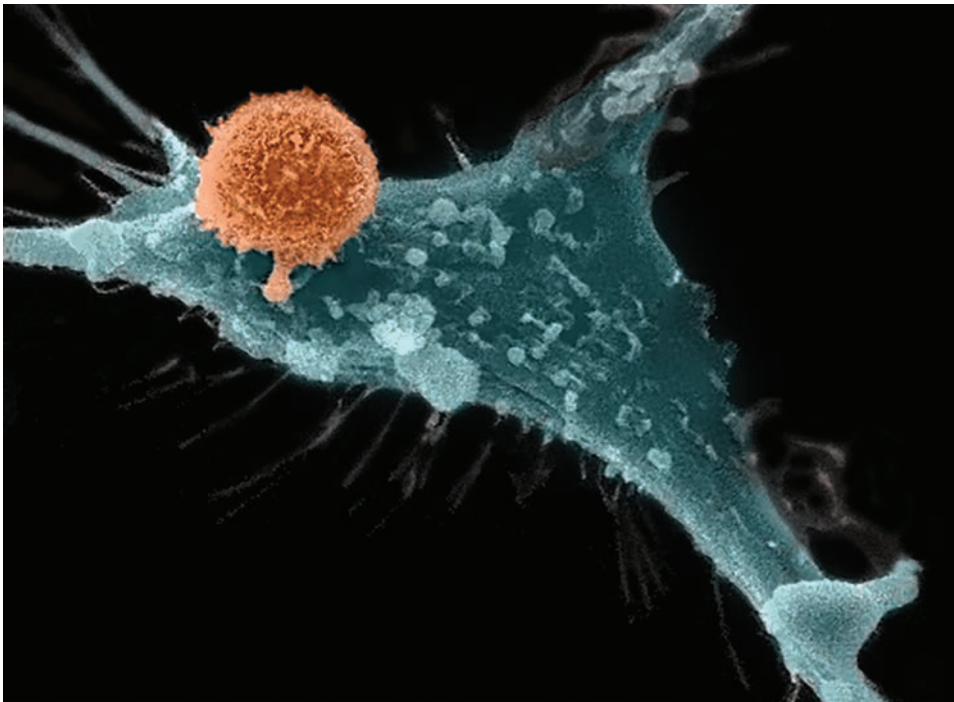
The Mass General Cellular Immunotherapy Program, directed by Dr. Maus, aims to generate a pipeline of genetically engineered CAR T cells to use as “living

drugs” in patients with cancer. The program is composed of a “research and discovery” arm, “a regulatory/translational” arm to test genetically-modified T cells in human subjects, and a “reverse translation” arm to examine the engraftment, persistence, and bioactivity of T cell products infused into patients. The immune profiling of patients is performed by the Immune Monitoring Laboratory, directed by Dr. Kathleen Gallagher.

Specifically, the engineered T cells that the Maus lab generates are intended to overcome specific obstacles observed in the clinic. The next generation T cells will:

1. *Contain molecular designs to enhance specificity, potency, and safety.*

Novel types of antigen receptors are in development to target multiple antigens on tumor cells, which improves elimination of heterogeneous tumor cells and prevents antigen-negative relapse while also decreasing the risk of targeting healthy cells. We are also using novel techniques to improve CAR T cell safety by regulating their activation and the molecules they release when activated. In liquid tumors, the focus is on improving the safety of CAR T cells, while in solid tumors, the focus is increasing their potency.



CAR-T Cell Targeting a Glioblastoma Cell Expressing EGFRvIII, Scanning Electron Micrograph; Credit: Bryan D. Choi, Mark B. Leick, and Marcela V. Maus.

2. Be administered in combination with other drugs to sensitize tumors to T cell mediated killing and/or potentiate T cell function.

Many of the small molecule drugs and antibodies used in the clinic exert their effects on signaling pathways in tumor cells, T cells, and other immune cells. We aim to discover synergistic drug/T cell combinations to increase safety and efficacy, and use genetic engineering tools to confer specific drug sensitivity, resistance, or enhanced molecular switches.

3. Have additional modifications that make CAR T cells (a) resistant to inhibitory mechanisms, (b) imageable, or (c) more feasible to manufacture and administer.

Control of T cell function is a complex process orchestrated by a variety of molecules, some of which deliver inhibitory signals. Tumors often express ligands or cytokines to inhibit T cell function. Using a single vector, genetically modified T cells can be

re-directed not only to recognize a new antigen on tumor cells, but also to be resistant to the inhibitory tumor microenvironment.

4. Further build on the basic biology and mechanisms that drive natural and engineered T cell functions.

We aim to understand the signaling mechanisms and effector functions used by CAR T cells versus native T cells, to further improve CAR T cell efficacy and safety. By understanding how CAR T cells kill tumor cells, we can also decipher how tumors cells become or are intrinsically resistant to killing by CAR T cells. We can then better engineer CAR T cells to prevent resistance from occurring.

Selected Publications:

Jan M, Scarfò I, Larson RC, Walker A, Schmidts A, Guirguis AA, Gasser JA, Slabicki M, Bouffard AA, Castano AP, Kann MC, Cabral ML, Tepper A, Grinshpun DE, Sperling AS, Kyung T, Sievers QL, Birnbaum ME, **Maus MV**, Ebert BL. Reversible ON- and OFF-switch chimeric antigen receptors controlled by lenalidomide. *Sci Transl Med*. 2021 Jan 6;13(575):eabb6295.

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Frigault MJ, Dietrich J, Martinez-Lage M, Leick M, Choi BD, DeFilipp Z, Chen YB, Abramson J, Crombie J, Armand P, Nayak L, Panzini C, Riley LS, Gallagher K, **Maus MV**. Tisagenlecleucel CAR T-cell therapy in secondary CNS lymphoma. *Blood*. 2019 Sep 12;134(11):860-866.